

POSTER PRESENTATION

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The 4-D landscape of the inflammatory response

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Background

It is widely accepted that chromatin 'responds' to physiological cues via protein:DNA interactions and nucleosome rearrangement [1,2], and that transcription plays a key role in its higher-order organization [3]. What remains elusive is how the nuclear landscape reshapes, in 3-D space and time, to facilitate such responses to unfold.

Materials and methods

We add tumour necrosis factor α (TNF α) to primary human endothelial cells and induce the inflammatory cascade; this is orchestrated by the transcription factor NF- κ B [4]. We monitor the response for 0-85 min postinduction using ChIP nucleosome-positioning studies, and chromosome conformation capture, all coupled to nextgeneration sequencing. We also apply a new approach, where the isolation of 'transcription factories' [5] is followed by RNA-seq to uncover nascent transcriptomes.

Results

First, we redefine early, intermediate, late, and oscillating TNFα-responsive genes, based on changing levels of nascent RNA. We then examine how these co-associate in specialized 'factories', some of which further specialize in transcribing responsive non-coding genes [6]. Contacts are driven by NF- κ B, and evolve as genes are differentially turned on and off over time. We also monitor nucleosome rearrangements genome-wide; these correlate with poised promoters before induction, and with nucleosome depletion as a result of transcriptional activation, NF- κ B binding, enhancer activity in TNFα-stimulated chromosomal domains.

Conclusions

We provide evidence for a prompt, within <30 min, reshaping of the genome in response to inflammation. This entails *de novo* associations of co-regulated coding and non-coding sequences in specialized 3-D networks that evolve over time, as well as extensive nucleosome depletion. We expect all extracellular cues to signal through analogous specialized networks and reassess our parsimonious model [7] for transcriptional regulation accordingly.

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References

- Grøntved L, Hager GL: Impact of chromatin structure on PR signaling: transition from local to global analysis. Mol Cell Endocrinol 2012, **357**·30-36
- Turner BM: The adjustable nucleosome: an epigenetic signaling module. Trends Genet 2012, 28:436-444.
- Papantonis A, Cook PR: Genome architecture and the role of transcription. Curr Opin Cell Biol 2010, 22:271-276.
- Smale ST: Hierarchies of NF-KB target-gene regulation. Nat Immunol 2011,
- Melnik S, Deng B, Papantonis A, Baboo S, Carr IM, Cook PR: The proteomes of transcription factories containing RNA polymerases I, II or III. Nat Methods 2011, 8:963-968.
- Papantonis A, Kohro T, Baboo S, Larkin JD, Deng B, Short P, Tsutsumi S, Taylor S, Kanki Y, Kobayashi M, Li G, Poh HM, Ruan X, Aburatani H, Ruan Y, Kodama T, Wada Y, Cook PR: TNFα signals through specialized factories where responsive coding and miRNA genes are transcribed. EMBO J 2012. 31:4404-4414.
- Kolovos P, Knoch TA, Grosveld FG, Cook PR, Papantonis A: Enhancers and silencers: an integrated and simple model for their function. Fniaenetics & Chromatin 2012, 5:1.

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