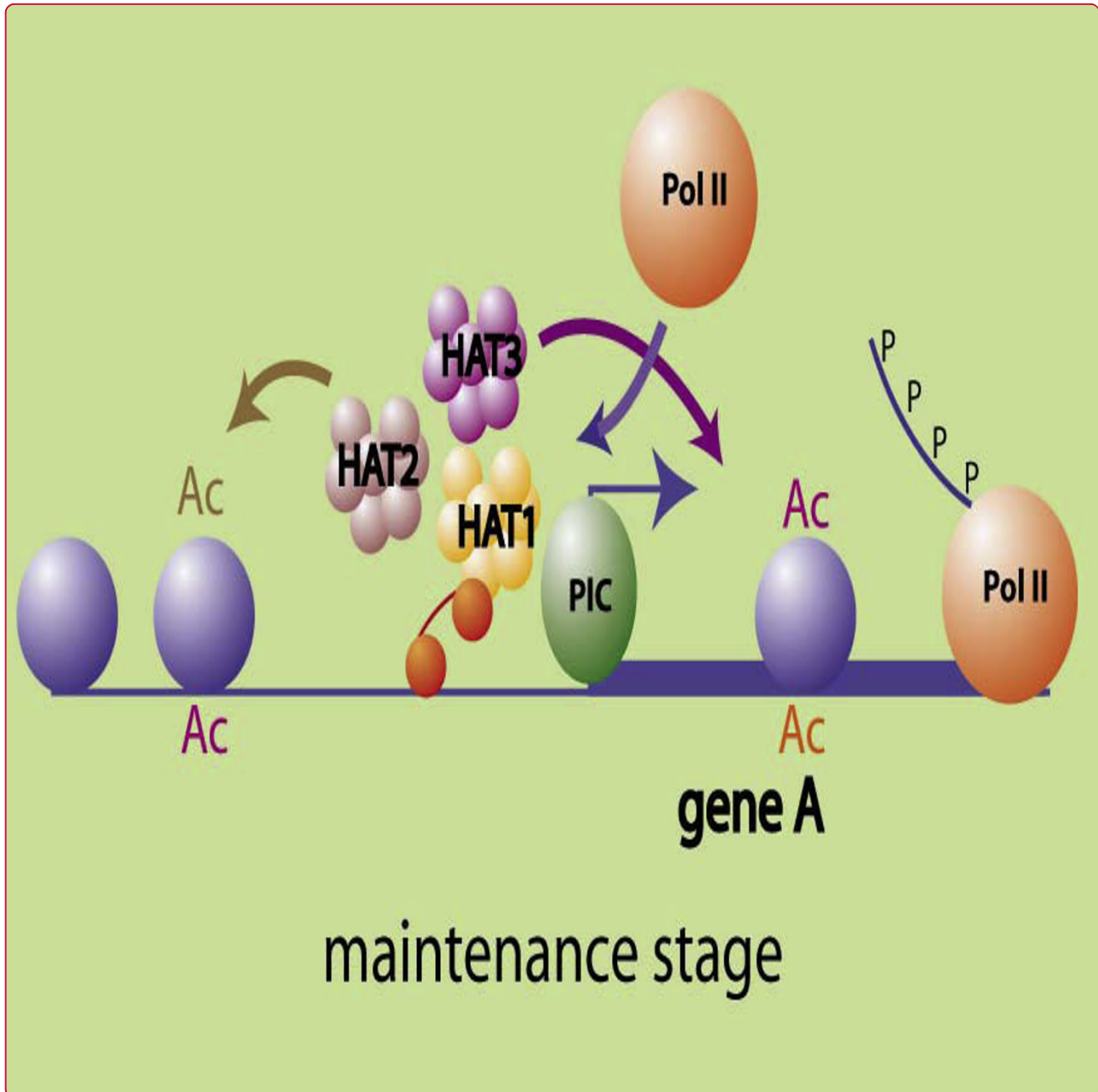


# Epigenetics & Chromatin



Lessons from genome-wide studies: an integrated definition of the coactivator function of histone acetyl transferases

Anamika *et al.*

REVIEW

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# Lessons from genome-wide studies: an integrated definition of the coactivator function of histone acetyl transferases

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## Abstract

Histone acetylation is one of the key regulatory mechanisms controlling transcriptional activity in eukaryotic cells. In higher eukaryotes, a number of nuclear histone acetyltransferase (HAT) enzymes have been identified, most of which are part of a large multisubunit complex. This diversity, combined with the large number of potentially acetylatable lysines on histones, suggested the existence of a specific regulatory mechanism based on the substrate specificity of HATs. Over the past decade, intensive characterisations of the HAT complexes have been carried out. However, the precise mode of action of HATs, and particularly the functional differences amongst these complexes, remains elusive. Here we review current insights into the functional role of HATs, focusing on the specificity of their action. Studies based on biochemical as well as genetic approaches suggested that HATs exert a high degree of specificity in their acetylation spectra and in the cellular processes they regulate. However, a different view emerged recently from genomic approaches that provided genome-wide maps of HAT recruitments. The careful analysis of genomic data suggests that all HAT complexes would be simultaneously recruited to a similar set of loci in the genome, arguing for a low specificity in their function. In this review, we discuss the significance of these apparent contradictions and suggest a new model that integrates biochemical, genetic and genome-wide data to better describe the functional specificity of HAT complexes.

## Introduction

Histone post-translational modifications have shown to be key regulators among transcription regulation mechanisms [1,2]. Histone acetylation is known to play an important role in the regulation of transcriptional activity in eukaryotic cells [3] by affecting higher-order folding of chromatin fibres, loosening of the contacts between the DNA and the nucleosomes and/or histone-nucleosome protein interactions [4-8]. Histone acetylation on various target lysines is in general positively associated with gene expression. Thus, HATs (also called lysine acetyl transferases or KATs) are thought to increase the decompaction of chromatin, which in turn may increase the accessibility of factors that promote

transcription [8-11]. In higher eukaryotes, two enzymatic families (GNAT and MYST), each containing a dozen histone acetyltransferase (HAT) enzymes, have been identified and have often been shown to be subunits of larger transcriptional coactivator complexes.

Over the past decade, two approaches were mainly used to better understand the functional specificity of HATs. First, *in vitro* acetylation assays were carried out to investigate the substrate specificity of distinct HATs. These analyses showed that HATs exert a certain degree of specificity for particular lysine residues on different histone tails. Second, *in vivo* gene inactivation studies allowed testing the HAT specificity by observing phenotypical effects caused by ablation of a particular HAT. Interestingly, most of these studies argued for a high degree of specificity in the developmental or gene expression phenotypes. More recently, availability of new high-throughput technologies such as chromatin immunoprecipitation sequencing (ChIP-seq) allowed the investigation of the recruitment of HATs and the deposition of acetylation marks at a

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genome-wide scale [12,13]. Contrary to the biochemical and genetic evidence, when carefully analysed, the genome-wide data suggest a low specificity in the recruitment and activity of HATs.

Here we comparatively review the conclusions of the above-mentioned three different approaches. Additionally, we discuss the significance of the conclusions made from each approach and try to reconcile new genome-wide evidence with existing knowledge in the form of a new model for the mode of action of HATs in transcriptional activation.

**Biochemistry: Each HAT has a specific histone acetylation spectrum modulated by its macromolecular complex**

The two aspects that were intensively investigated immediately after the discovery of HATs were (1) the identification of the protein complexes in which the individual HAT enzyme is incorporated and (2) the determination of the acetylation spectrum associated with each HAT individually and then in the corresponding complexes (both reviewed in Figure 1 for mammalian cells). Several conclusions emerged from these studies. First, it was shown that most of the HAT enzymes are part of large multisubunit complexes. For example, Tip60 (Tat-interactive protein 60 kD; also called KAT5) is contained in the 18 subunits of the NuA4 complex [14], whereas GCN5 (general control of amino-acid synthesis 5; also called KAT2A) or PCAF (p300/CBP-associated protein; also called KAT2B) are members of the 19 subunits of the SAGA complex and the 10 subunits of the ATAC complex [15]. Second,

several studies described how the association of the HAT enzymes with their partners in the corresponding complexes can modify the specificity of the HATs on histones (i.e., *Drosophila* GCN5 was shown to have different targeting specificity in ATAC or SAGA complexes [16]). Third, each HAT acetylates a defined spectrum of target lysines, and these acetylation sites only partially overlap between individual HAT complexes (see Figure 1 for review in mammalian systems). Taken together, these observations suggest a high specificity of each HAT when incorporated in its corresponding macromolecular complex, suggesting that this integration is required for the full and specific activity *in vivo*. Moreover, these studies suggest that each HAT complex should create a specific signature on its target loci because a distinguishable acetylation pattern is observed for each complex tested *in vitro* [11,17].

**Genetics: HATs exert a high degree of functional specificity**

Another aspect that was systematically addressed to better understand the function of HATs is the phenotypical analysis of the effect of distinct ablation of HATs *in vivo* (reviewed in Figure 2). Genetic knockout (KO) studies of HATs during mouse development reveal a significant variability in the phenotypes of HAT ablations. For example, *Tip60*<sup>-/-</sup> embryos do not develop beyond blastocyst stage (i.e., they die at embryonic day (ED) 3.5) [18], while *Gcn5*<sup>-/-</sup> embryos die at around ED 10.5 [19]. Moreover, the maintenance of embryonic stem cell (ESC) pluripotency can be used as a second phenotypical readout.

	molecular complex	HAT Family	H3				H4				H2A		H2B				references	
			9	14	18	23	5	8	12	16	5	9	5	12	15	20		
GCN5 - KAT2A/ PCAF - KAT2B	SAGA	GNAT	■	■	■	■												[15, 27],
	ATAC		■	■			■	■	■	■								[15, 28, 29]
	free		■	■														
MOF - KAT8	MSL	MYST								■								[18]
	NSL						■	■										[18]
	free		ND	ND	ND	ND					ND	ND	ND	ND	ND	ND	ND	[18]
TIP60 - KAT5	NuA4	MYST	■	■	■	■	■	■	■	■	■							[30-32]
	free		■	■	■	■	■	■	■	■	■	■						[32, 33]
p300 - KAT3B	p300/CBP	p300/CBP	ND	■	■	ND	■	■	■	ND	■	ND	■	■	■	■	ND	[34]
CBP - KAT3A	p300/CBP	p300/CBP	ND	■	■	ND	■	■	■	ND	■	ND	■	■	■	■	ND	[34]

**Figure 1 Substrate specificity of histone acetyltransferases (HATs) *in vitro*.** Review of the *in vitro* substrate specificity described in mammalian systems for the studied five HAT enzymes, either as a free protein or within their respective macromolecular complexes. Dark grey boxes represent lysine residues highly acetylated, light grey boxes represent residues where weak acetylation activity has been observed, and white boxes represent residues where no acetylation is detected. ND represents residues that have not been tested for acetylation. References [15,18,27-34] cited in figure.

	molecular complex	HAT family	Phenotypic readout		
			mouse knock out	mESC pluripotency	references
GCN5 - KAT2A	SAGA/ATAC	GNAT	ED 10.5	-	[19, 20]
PCAF - KAT2B			viable	-	[19, 21]
MOF - KAT8	MSL/NSL	MYST	ED 3.5	+	[21, 35]
TIP60 - KAT5	NuA4		ED 3.5	+	[21, 36]
CBP - KAT3A	p300/CBP	p300/CBP	ED 11.5	ND	[22]
p300 - KAT3B	p300/CBP		ED 11.5	-	[21, 22]

**Figure 2 Phenotypes observed in gene disruption studies of HATs.** Phenotypes associated with HAT genetic knockout (KO) or RNAi targeting in mouse development and embryonic stem cell (ESC) pluripotency, respectively. For mouse KO, the day of embryonic death (ED) for homozygous animals is presented as a phenotypic readout parameter. For the effect on mouse embryonic stem cell (mESC) pluripotency, any observation (e.g., flattening of the cells, differentiation) was simplified as a positive phenotype (+). ND represents HATs for which the parameter was not determined. References [19-22,35,36] cited in figure.

Using RNAi knockdown of Tip60, this enzyme was shown to be required for ESC pluripotency, while GCN5 or PCAF seems to be dispensable (Figure 2) [20,21]. Together, these results suggest that each HAT has key roles at different stages of development that seem to be independent of and distinct from other HATs.

A remarkable feature emerging from sequence alignment of vertebrate HATs is the presence of highly related paralogs [i.e., CBP (CREB-binding protein, KAT3A)/p300 (KAT3B) or Gcn5/PCAF], whereas for most of the nuclear HATs only a single gene is present in nonvertebrate species. This observation raised the question of the functional importance of closely related paralogs in vertebrates. Answers were partially obtained by crossing mice carrying individual homozygous or heterozygous KOs of the given paralogs and comparing double KOs to that of the single KOs. For example, while the single CBP or p300 heterozygous mutants exert a milder phenotype than the homozygous mutants, the mice heterozygous for both HATs show phenotypes similar to the homozygous depletion of one or the other paralog [22]. This result suggests that there is a functional redundancy between these paralogs and that in certain cases the dose of expression of the two paralogs is crucial for the proper development of the animals.

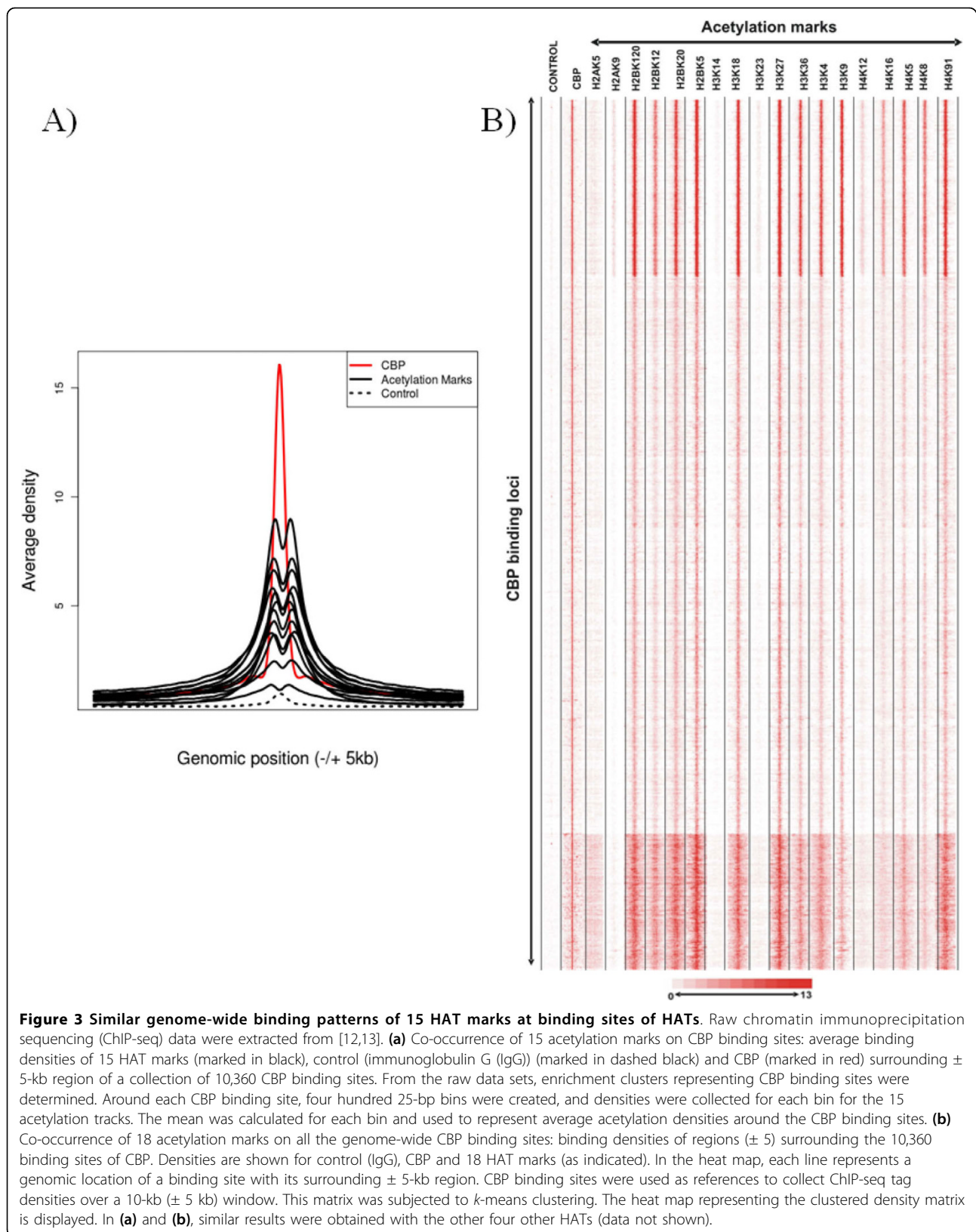
Taken together, these studies suggest that, amongst HATs, a high degree of functional specificity exists, with the exception of closely related paralogs that rather reflect the importance of gene expression dosage than functional specificity.

#### Genome-wide mapping: HATs are co-occurring at high frequency, creating hyperacetylated environment

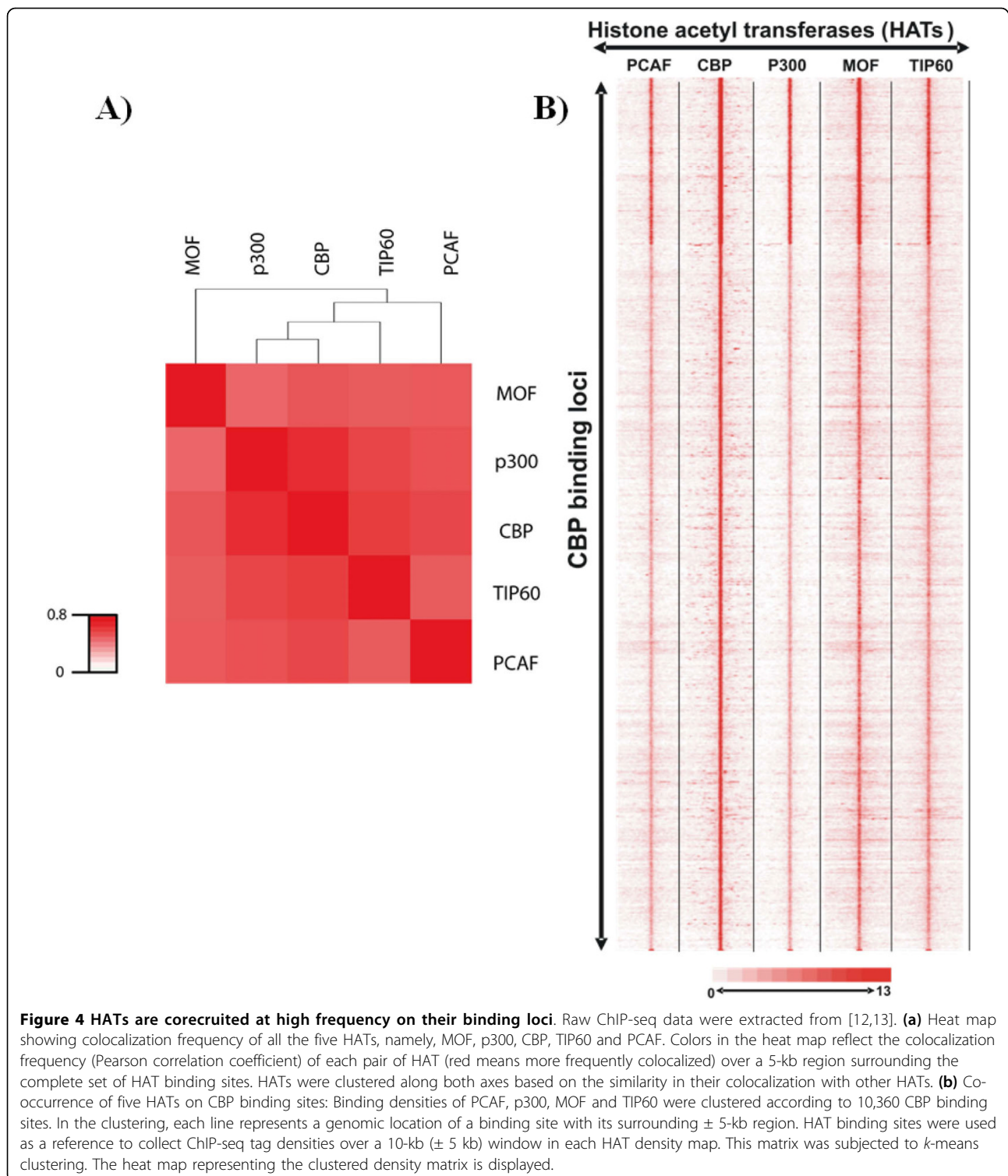
In the past few years, our understanding of genome regulation has tremendously progressed due to the introduction of ChIP combined with microarray (ChIP-chip)

or with high-throughput sequencing (ChIP-Seq). These experiments have allowed us to analyse the presence of a particular genomic feature at the genome-wide scale. Recently, several systematic ChIP-seq studies have been published, providing genome-wide mapping data for HATs and acetylation marks in resting human T cells [12,13].

This resource has given us the opportunity to analyse the specificity of genome-wide HAT recruitments. We have first extracted genome-wide distributions of five HATs [namely, p300, CBP, MOF (males absent on the first; KAT8), PCAF, and Tip60) and 18 acetylation marks and then systematically analysed the presence of each of the 18 acetylation marks at the binding sites of the five distinct HATs. Interestingly, this analysis revealed that, when comparing the genome-wide binding sites of a given HAT with that of the 18 different histone acetylation marks, no specific or unique acetylation pattern for any of the five HATs could be identified (Figures 3a and 3b; shown for CBP). In each of these “HAT-acetylation mark” comparisons, variations in acetylation levels between different categories of loci can be observed, but with the same patterns for all the residues tested (except for histone H2AK5ac, H3K14ac and H3K23ac for which the antibodies used in the ChIP may have been too weak for the analysis). The most likely hypothesis to explain these results is that HATs are co-occurring at these sites. To further analyse this possibility, the binding specificity of a given HAT versus the four other HATs has been tested. Interestingly, this analysis revealed that each genomic locus bound by a given HAT is also bound to some extent by the four other HATs (Figures 4a and 4b). It appears that the binding of each HAT correlates with the binding of all the other HATs tested (Figure 4a; Pearson correlation > 0.6).







The detected co-occurrence of HATs and the consequent broad acetylation patterns observed can suggest two different scenarios. First, since in ChIP cell populations rather than a single cell are analysed, we cannot exclude that the recruited HATs could occur in a

stochastic manner. This would imply that in a given cell, one HAT is recruited to a particular locus, while in another cell, a distinct HAT is recruited instead to the same locus. In this scenario, the observed lack of histone acetylation specificity would come from the lack of

specificity in the mechanisms that recruit HATs to particular loci. In this case, the acetylation specificity would not be an important feature for regulation, but only the acetylation *per se* would be requested for proper activation.

In the second scenario, HATs would work collaboratively. In all cells, all the studied HATs would be permanently recruited and released at every bound loci in a dynamic fashion. This scenario implies also that many transcriptional coactivators can be dynamically recruited all the time to a set of regulated loci to modify (acetylate) the given chromatin environment. Such a mechanism for coactivator action has already been suggested by Hager *et al.* [23]. Thus, gene regulation would mainly rely on the local abundance of the different coactivators rather than on the recruitment of a specific coactivator and its corresponding specific pattern of histone modifications. In this case, there would be a collaborative effect of all the HATs on the deposition of histone acetylation marks at the regulated set of genes.

#### **An integrated model depicting mode of action of HATs**

Taken together, the evidence emerging from genomics, biochemical and genetic analyses of HAT action result in apparently contradictory conclusions. While both the biochemistry and the genetic data argue for a rather high degree of specificity in the functional spectrum of HATs (Figure 5a), the current genome-wide evidence suggests that HATs are in general redundantly recruited to the same set of loci (Figure 5b), suggesting low specificity in the action of HATs. To reconcile the different observations, we first confront each line of evidence and then propose a new model that may better explain the *in vivo* action of HAT complexes (Figure 5c).

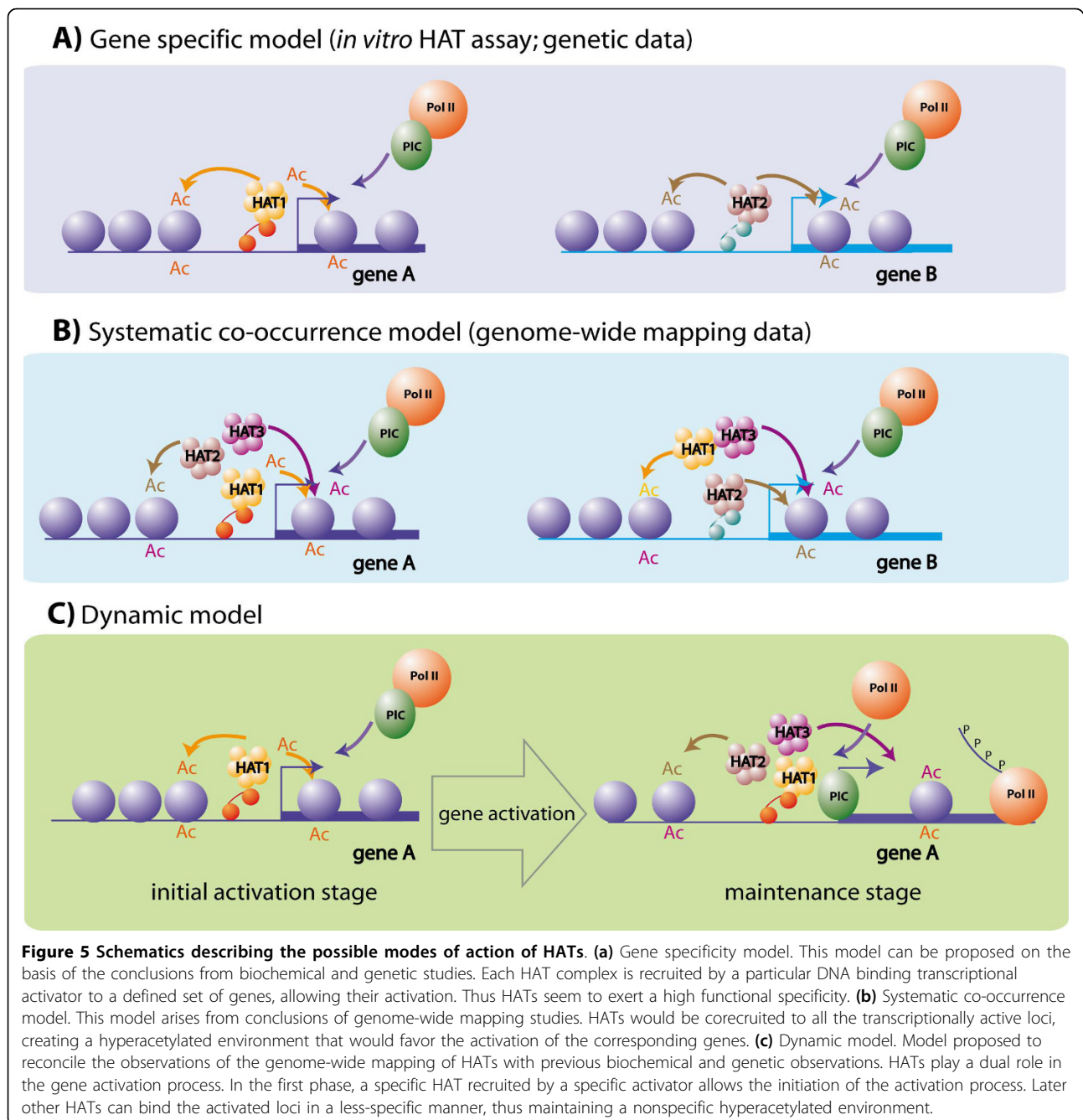
The genome-wide results do not directly contradict the biochemical evidence, since the broad *in vivo* co-occurrence of HAT enzymes at many genomic loci would explain the observation of multily sine hyperacetylated loci *in vivo*. However, this model is challenged by the conclusions of the genetic studies. It is clear from the KO studies that different HATs cannot compensate for each other's function. Moreover, if each process would require all the HATs, one would expect that all HAT KOs would result in the same phenotype, since all HAT-dependent processes should be affected. Thus a model assuming a complete overlap in HAT function can be excluded, and new hypotheses have to be raised to integrate the new lines of genome-wide evidence.

The time is a variable that was not introduced in the previous formulation of hypotheses. The analysed ChIP-seq data were all produced at a time  $t$  that reflects a single stage of the transcription activation process. Since the analysed genome-wide data were generated in resting somatic cells (CD4+ T cells) that are not challenged, one

can assume that they exert only "routine" gene expression programs compared to the major transcriptional changes happening during differentiation of a tissue or an organism. We hypothesize that the genome-wide data obtained in the resting somatic cells reflect a "maintenance stage" in the gene activation process (Figure 5c). In our novel model, we propose that HATs could have a dual mode of action by distinguishing "initiation" and "maintenance" stages during the transcription activation process. The "initiation" stage would reflect the initial steps in the switch from an inactive to active transcription state, while the "maintenance" stage would represent the stabilisation of an active transcription stage over the time. In the initiation stage, the transcriptional activation of a given gene would be highly dependent on the specific recruitment of one given HAT (Figure 5c, left). Following this "initiation" stage, through a sequential process, this initial activation would lead to the consequent recruitment or binding of multiple HATs, which may recognize the initially open and acetylated environment in a less specific manner. This hypothesis is in good agreement with the findings that many HAT-containing complexes contain subunits with bromodomains that are thought to bind to acetylated histone tails. The binding of several HATs to the initially acetylated locus would thus serve to maintain the acetylation level and by consequence the activation state of a given locus (Figure 5c, right).

Our model would thus explain why in dynamic developmental processes HATs show high functional specificity (genetic data) that may not be the case in resting differentiated somatic cells. During cell differentiation, the high complexity level of transcriptional regulatory networks is a prerequisite for the orchestration of the time-controlled initiation of the proper transcriptional programs. However, in resting somatic tissues, where the number of newly initiated transcription processes is likely to be lower than during cell commitment, the large panel of activators would redundantly maintain activation states. This could explain why high redundancy in the recruitment of HATs is observed in resting cells, while their functional specificity is needed to achieve proper embryo development.

The above-presented data [12] is focused on catalytic subunits of HAT complexes. However, as discussed previously, a given HAT enzyme is (with the notable exception of CBP/p300) often embedded in more than one large multisubunit complex. Therefore, although the distinct HAT enzymes appear to be corecruited at many loci, they can represent the occurrence of different HAT complexes. Indeed, two recent studies analysed the differences of the recruitment between the two MOF-containing complexes (called NLS and MLS) at the genome-wide scale in *Drosophila* [24,25]. The studies, by



targeting NLS- or MLS-specific subunits, show a high specificity in the recruitment of these complexes. Thus, by studying individually the well-defined HAT complexes, the resolution of the analysis will increase, leading to a more precise separation of the observed overlap in the recruitment of the HATs. Nevertheless, the existence of specific loci, where a single HAT is recruited should not be excluded. However, according to the analysis of the data from the Zhao lab [12], these events appear to be the exception rather than the rule.

The very recent improvement of the genome-wide mapping technologies and the development of novel ChIP-grade antibodies against HATs and complex-specific subunits will allow researchers to verify these genome-wide observations and provide new insights into the functional roles of HATs. For example, the time course analysis of different HAT recruitments upon dynamic activation of a particular gene networks, for example, *in vitro* stem cell differentiation systems [26], should allow researchers to test the proposed model.



## Conclusions

HAT-containing complexes are key components of chromatin-mediated transcriptional regulatory networks. Proper understanding and modelling of their mode of action and function within these networks is a prerequisite for accurate prediction of the behaviour of transcriptional systems. Recent technology breakthrough in the postgenomic area allowed new insights into the function of HATs, but were often interpreted with the liability of ignoring previously available biochemical and genetic data, leading to oversimplified models. Here we propose novel considerations that could reconcile the different lines of evidence in a unified model that describes the mode of action of HATs more precisely. Our current model proposes that HATs are differently required, depending on the stage of gene activation, with a high functional specificity in the early gene activation stage and with a less-specific functionality in the later maintenance stage.

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## Authors' contributions

ARK, KA and LT contributed to the study design. KA and ARK performed the bioinformatics analyses. KA, ARK, DD, JT, OP and LT wrote the manuscript. All authors read and approved the final manuscript.

## Competing interests

The authors declare that they have no competing interests.

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## References

1. Kingston RE, Bunker CA, Imbalzano AN: **Repression and activation by multiprotein complexes that alter chromatin structure.** *Genes Dev* 1996, **10**:905-920.
2. Eberharter A, Becker PB: **Histone acetylation: a switch between repressive and permissive chromatin: second in review series on chromatin dynamics.** *EMBO Rep* 2002, **3**:224-229.
3. Shahbazian MD, Grunstein M: **Functions of site-specific histone acetylation and deacetylation.** *Annu Rev Biochem* 2007, **76**:75-100.
4. Tse C, Sera T, Wolffe AP, Hansen JC: **Disruption of higher-order folding by core histone acetylation dramatically enhances transcription of nucleosomal arrays by RNA polymerase III.** *Mol Cell Biol* 1998, **18**:4629-4638.
5. Carruthers LM, Hansen JC: **The core histone N termini function independently of linker histones during chromatin condensation.** *J Biol Chem* 2000, **275**:37285-37290.
6. Wolffe AP, Hansen JC: **Nuclear visions: functional flexibility from structural instability.** *Cell* 2001, **104**:631-634.
7. Fischle W, Wang Y, Allis CD: **Histone and chromatin cross-talk.** *Curr Opin Cell Biol* 2003, **15**:172-183.
8. Shogren-Knaak M, Ishii H, Sun JM, Pazin MJ, Davie JR, Peterson CL: **Histone H4-K16 acetylation controls chromatin structure and protein interactions.** *Science* 2006, **311**:844-847.
9. Akhtar A, Becker PB: **Activation of transcription through histone H4 acetylation by MOF, an acetyltransferase essential for dosage compensation in *Drosophila*.** *Mol Cell* 2000, **5**:367-375.
10. Krajewski WA, Becker PB: **Reconstitution of hyperacetylated, DNase I-sensitive chromatin characterized by high conformational flexibility of nucleosomal DNA.** *Proc Natl Acad Sci USA* 1998, **95**:1540-1545.
11. Sterner DE, Berger SL: **Acetylation of histones and transcription-related factors.** *Microbiol Mol Biol Rev* 2000, **64**:435-459.
12. Wang Z, Zang C, Cui K, Schones DE, Barski A, Peng W, Zhao K: **Genome-wide mapping of HATs and HDACs reveals distinct functions in active and inactive genes.** *Cell* 2009, **138**:1019-1031.
13. Wang Z, Zang C, Rosenfeld JA, Schones DE, Barski A, Cuddapah S, Cui K, Roh TY, Peng W, Zhang MQ, Zhao K: **Combinatorial patterns of histone acetylations and methylations in the human genome.** *Nat Genet* 2008, **40**:897-903.
14. Sapountzi V, Logan IR, Robson CN: **Cellular functions of TIP60.** *Int J Biochem Cell Biol* 2006, **38**:1496-1509.
15. Nagy Z, Riss A, Fujiyama S, Krebs A, Orpinell M, Jansen P, Cohen A, Stunnenberg HG, Kato S, Tora L: **The metazoan ATAC and SAGA coactivator HAT complexes regulate different sets of inducible target genes.** *Cell Mol Life Sci* 2009.
16. Guelman S, Suganuma T, Florens L, Swanson SK, Kiesecker CL, Kusch T, Anderson S, Yates JR, Washburn MP, Abmayr SM, Workman JL: **Host cell factor and an uncharacterized SANT domain protein are stable components of ATAC, a novel dAda2A/dGcn5-containing histone acetyltransferase complex in *Drosophila*.** *Mol Cell Biol* 2006, **26**:871-882.
17. Brown CE, Lechner T, Howe L, Workman JL: **The many HATs of transcription coactivators.** *Trends Biochem Sci* 2000, **25**:15-19.
18. Cai Y, Jin J, Swanson SK, Cole MD, Choi SH, Florens L, Washburn MP, Conaway JW, Conaway RC: **Subunit composition and substrate specificity of a MOF-containing histone acetyltransferase distinct from the male-specific lethal (MSL) complex.** *J Biol Chem* 2010, **285**:4268-4272.
19. Xu W, Edmondson DG, Evrard YA, Wakamiya M, Behringer RR, Roth SY: **Loss of Gcn5l2 leads to increased apoptosis and mesodermal defects during mouse development.** *Nat Genet* 2000, **26**:229-232.
20. Ding L, Paszkowski-Rogacz M, Nitzsche A, Slabicki MM, Heninger AK, de Vries I, Kittler R, Junqueira M, Shevchenko A, Schulz H, Hubner N, Doss MX, Sachinidis A, Hescheler J, Iacone R, Anastasiadis K, Stewart AF, Pisabarro MT, Caldarelli A, Poser I, Theis M, Buchholz F: **A genome-scale RNAi screen for Oct4 modulators defines a role of the Paf1 complex for embryonic stem cell identity.** *Cell Stem Cell* 2009, **4**:403-415.
21. Fazio TG, Huff JT, Panning B: **An RNAi screen of chromatin proteins identifies Tip60-p400 as a regulator of embryonic stem cell identity.** *Cell* 2008, **134**:162-174.
22. Yao TP, Oh SP, Fuchs M, Zhou ND, Ch'ng LE, Newsome D, Bronson RT, Li E, Livingston DM, Eckner R: **Gene dosage-dependent embryonic development and proliferation defects in mice lacking the transcriptional integrator p300.** *Cell* 1998, **93**:361-372.
23. Hager GL, McNally JG, Misteli T: **Transcription dynamics.** *Mol Cell* 2009, **35**:741-753.
24. Raja SJ, Charapitsa I, Conrad T, Vaquerizas JM, Gebhardt P, Holz H, Kadlec J, Fraterman S, Luscombe NM, Akhtar A: **The nonspecific lethal complex is a transcriptional regulator in *Drosophila*.** *Mol Cell* 2010, **38**:827-841.
25. Prestel M, Feller C, Straub T, Mitlöhner H, Becker PB: **The activation potential of MOF is constrained for dosage compensation.** *Mol Cell* 2010, **38**:815-826.
26. Bibel M, Richter J, Lacroix E, Barde YA: **Generation of a defined and uniform population of CNS progenitors and neurons from mouse embryonic stem cells.** *Nat Protoc* 2007, **2**:1034-1043.
27. Brand M, Yamamoto K, Staub A, Tora L: **Identification of TATA-binding protein-free TAFII-containing complex subunits suggests a role in nucleosome acetylation and signal transduction.** *J Biol Chem* 1999, **274**:18285-18289.

28. Guelman S, Kozuka K, Mao Y, Pham V, Solloway MJ, Wang J, Wu J, Lill JR, Zha J: **The double-histone-acetyltransferase complex ATAC is essential for mammalian development.** *Mol Cell Biol* 2009, **29**:1176-1188.
29. Wang YL, Faiola F, Xu M, Pan S, Martinez E: **Human ATAC Is a GCN5/PCAF-containing acetylase complex with a novel NC2-like histone fold module that interacts with the TATA-binding protein.** *J Biol Chem* 2008, **283**:33808-33815.
30. Altaf M, Auger A, Monnet-Saksouk J, Brodeur J, Piquet S, Cramet M, Bouchard N, Lacoste N, Utley RT, Gaudreau L, Côté J: **NuA4-dependent acetylation of nucleosomal histone H4 and H2A directly stimulates incorporation of H2A.Z by the SWR1 complex.** *J Biol Chem* 2010, **285**:15966-15977.
31. Ikura T, Ogryzko VV, Grigoriev M, Groisman R, Wang J, Horikoshi M, Scully R, Qin J, Nakatani Y: **Involvement of the TIP60 histone acetylase complex in DNA repair and apoptosis.** *Cell* 2000, **102**:463-473.
32. Allard S, Utley RT, Savard J, Clarke A, Grant P, Brandl CJ, Pillus L, Workman JL, Cote J: **NuA4, an essential transcription adaptor/histone H4 acetyltransferase complex containing Esa1p and the ATM-related cofactor Tra1p.** *EMBO J* 1999, **18**:5108-5119.
33. Yamamoto T, Horikoshi M: **Novel substrate specificity of the histone acetyltransferase activity of HIV-1-Tat interactive protein Tip60.** *J Biol Chem* 1997, **272**:30595-30598.
34. Schiltz RL, Mizzen CA, Vassilev A, Cook RG, Allis CD, Nakatani Y: **Overlapping but distinct patterns of histone acetylation by the human coactivators p300 and PCAF within nucleosomal substrates.** *J Biol Chem* 1999, **274**:1189-1192.
35. Thomas T, Dixon MP, Kueh AJ, Voss AK: **Mof (MYST1 or KAT8) is essential for progression of embryonic development past the blastocyst stage and required for normal chromatin architecture.** *Mol Cell Biol* 2008, **28**:5093-5105.
36. Hu Y, Fisher JB, Koprowski S, McAllister D, Kim MS, Lough J: **Homozygous disruption of the Tip60 gene causes early embryonic lethality.** *Dev Dyn* 2009, **238**:2912-2921.

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